

# NTP/NIEHS CONTRACT NO.:N01-ES-75409

Pubertal Vinclozolin Study
Pubertal Methoxychlor Study
Pubertal Flutamide Study
Pubertal Ethinyl Estradiol Study
RACB 20103
RACB 20105
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# PUBERTAL TOXICITY STUDY OF VINCLOZOLIN AND FLUTAMIDE IN MALE SPRAGUE-DAWLEY RATS AND METHOXYCHLOR AND ETHINYL ESTRADIOL IN FEMALE SPRAGUE-DAWLEY RATS WHEN ADMINISTERED IN CORN OIL BY ORAL GAVAGE

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#### 1.0 INTRODUCTION

#### 1.1 Proposed Investigations/Rationale for Dose Selection

Numerous pharmaceutical and environmental agents have been shown to alter the timing of pubertal development in mammals since puberty is a time of rapid interactive endocrine and morphological changes. The purpose of the current study is to provide data on proposed procedures to evaluate the effects of Vinclozolin, Methoxychlor, Flutamide and Estradiol on pubertal development in the intact juvenile/peripubertal male and female rat. The proposed design will detect agents that have antithyroid, estrogenic, androgenic, antiandrogenic activity, or alter puberty via changes in follicle stimulating hormone (FSH), luteinizing hormone (LH), prolactin, growth hormone (GH), or hypothalamic function (Stoker *et al.*, 2000 and Goldman *et al.*, 2000).

Vinclozolin, or 3-(3,5-dichlorophenyl)-5-methyl-5-vinylox-azolidine-2,4-dione, is a fungicide used on fruits, vegetables, turfgrass, and ornamental plants (U.S. EPA, 1998). In vivo, vinclozolin inhibits AR-dependent gene expression (Kelce et al., 1997) and produces a spectrum of anatomical defects.

Administration of vinclozin (400 mg/kg) to rats on gestational day (GD) 14 through postnatal day (PND) 3 resulted in effects similar to those caused by flutamide, a well known AR antagonist. These effects include reduced anogenital distance (AGD); persistent nipples; cleft phalllus; hypospadias; reduces weights of the ventral prostate, seminal vesicles and epididymis; and reduces sperm count (Gray et al., 1994; 1999a). Exposing weanling male rats to the antiandrogenic pesticides p,p'- DDE or vinclozolin delays pubertal development in weanling male rats as indicated by delayed preputial separation and increases body weight at puberty. In contrast to delays associated with exposure to estrogenic substances, antiandrogens do not inhibit food consumption or retard growth (Anderson, et al., 1995b).

Flutamide (4'-nitro-3'-trifluoromethyl-isobutyranilide) is a potent nonsteroidal androgen receptor antagonist that has been used therapeutically to treat androgen-dependent prostrate cancer (Delaere and Van Thillo, 1991;Murphy *et al.*, 1991) and as a tool to study male reproductive development. Studies in rats have demonstrated that pre- or postnatal flutamide (6.25 to 50 mg/kg) exposure alters androgen-dependent reporductive development (Imperato-McGinley *et al.*, 1992; Kassim *et al.*, 1997) and has been shown to produce decreased reproductive organ weights, feminization of male external genitalia, altered

androgen-dependent testicular descent, and retention of nipples when male offspring are exposed *in utero* (Imperato-McGinley *et al.*, 1992).

Methoxychlor has been used for nearly 50 years for insect and larval control. Its advantages over DDT are that methoxychlor is more readily metabolized and excreted by mammalian systems (Kapoor *et al.*, 1970), and therefore there is less bioconcentration than with DDT. This metabolism also yields monoand bis-hydroxy metabolites of methoxychlor (Bulger et al., 1978), which helps explain both the uterotrophic effects noted earlier for methoxychlor (Tullner, 1961) and the observations that methoxychlor *in vivo* reduced the uterine uptake of radiolabeled estradiol (Welch *et al.*, 1969). Treatment with methoxychlor at 5, 50, or 150 mg/kg for the week before and the week after birth to PND 7 resulted in unchanged anogenital distance, accelerated vaginal opening, delayed prepuce separation. Methoxychlor at 50 and 150 mg/kg disrupted adult estrous cyclicity and reduced epididymal sperm counts and testis weights

As cited by Goodman and Gilman's The Pharmacological Basis of Therapeutics, (1996), estrogens are among the most commonly prescribed drugs in the United States. The two major uses are as components of combination oral contraceptives and hormone replacement therapy in postmenopausal women. The pharmacological considerations for estrogen use in oral contraceptives and post menopaousal hormone replacement are substantially different, primarily because of the doses used. Historically, conjugated estrogens have been the most common agents for postmenopausal use, and 0.625 mg/kg/day is effective in most women. In contrast, most combination oral contraceptives in current use employ 20 to 35 ug/day of ethinyl estradiol. Conjugated estrogens and ethinyl estradiol differ widely in their oral potencies; for example, a dose of 0.625 mg of conjugated estrogens generally is considered equivalent to 5 to 10 ug of ethinyl estradiol.

Several authors have demonstrated the estrogenic responses to ethinyl estradiol in rodents. Laws et al. (2000) showed that *in vivo* studies indicated the 3-day uterotrophic assay in prepubertal rats was best for detecting estrogenic activity when compared with all other end points, based upon the dose-response data for ethinyl estradiol (0.01-0.1 mg/kg, oral), 4-tert-octylphenol (50-200 mg/kg, oral), and 4-nonylphenol (25-100 mg/kg, oral). Although oral doses of ethinyl estradiol (0.01 mg/kg) induced a significant increase in uterine weight in the prepubertal rat, this dose was ineffective for stimulating a

similar response in ovariectomized adult rats. The age of vaginal opening was advanced following oral exposure from postnatal days 21-35 to ethinyl estradiol (0.01 mg/kg). Ethinyl estradiol advanced the day of vaginal opening by  $6.0 \pm 0.18$  days (30.6 days in control vs. 24.6 days in treated). In addition, the number of 4-5 day estrous cycles was reduced during a 25 day exposure to ethinyl estradiol (0.01 mg/kg).

Advanced vaginal opening was also demonstrated by Odum et al., (1997) using doses of 2-400 ug/kg/day, subcutaneous, and Singh and Kamgoj, (1980) using doses of 5 ug/kg/day for 5 days. Singh and Kamboj (1980) also showed an advance in cornified vaginal cells.

The data in this study should result in advanced vaginal opening, advanced first estrous and onset of estrous cycles, and/or persistent vaginal estrus in the ethinyl estradiol and methoxychlor females exposed *in utero* and in delayed preputial separation, decreased reproductive organ weights, altered external genitalia, and/or retention of nipples in the vinclozolin and flutamide males exposed *in utero*.

# 1.2 Regulatory Compliance

This study will be conducted according to a modification of the Health Effects Test Guidelines

OPPTS 870.8300 Reproduction and Fertility Effects and in compliance with the Food and Drug

Administration Good Laboratory Practice Regulations for Nonclinical Laboratory Studies (1987). All

procedures will follow TherImmune Standard Operating Procedures.

#### 1.3 Quality Assurance

The protocol, in-life phases, data, and the final report will be audited by TherImmune Quality

Assurance. Critical phases to be audited for each generation will be selected by the Director of Quality

Assurance.

# 1.4 Testing Facility

TherImmune Research Corporation (TherImmune)

15 Firstfield Road

Gaithersburg, MD 20878

#### 2.0 TEST ARTICLE

#### 2.1 Characterization of Test Articles

# 2.1.1 Vinclozolin (from MSDS)

Identity: Vinclozolin

R.O.W. ID No.: 1317B

Source: Battelle Organic Synthesis Group

CAS No.: 50471-44-8

Lot No.: 102996

Molecular Wt: 286.1g/mol

Formula:  $C_{12}H_9NO_3CL_2$ 

C1 H 6 CH4

Purity: 99.6%

Storage:

Structure:

Test Article: Store at room temperature (~25°C) and protected from light.

Formulation: Stored in sealed amber glass bottles at room temperature (~25°C) and

protected from light

Stability:

Test Article: Analyze every  $24 \pm 2$  weeks to verify stability.

Formulation: Dose formulations (2 mg/mL) are stable for 42 days at temperatures of

25°C, 5°C, and -20°C in sealed amber glass bottles and protected from

light.

# 2.1.2 Methoxychlor (from MSDS)

Identity: Methoxychlor

R.O.W. ID No.: 1321B

Source: Sigma Chemical Co.

CAS No.: 72-43-5

Lot No.: 124F0575

Molecular Wt: 345.7

Formula:  $C_{16}H_{15}Cl_3O_2$ 

Structure: CH<sub>9</sub>O CH<sub>9</sub>OCH<sub>9</sub>

Purity: 95%

Storage:

Test Article: Stored in a sealed container under nitrogen and protected from light at

ambient temperature (23 to 28°C)

Formulation: Stored in sealed glass vials at refrigerator temperature.

Stability:

Test Article: Analyze every 24± 2 weeks to verify stability.

Formulation: Dose formulations (1.82 mg/mL in corn oil ) are stable for 30 days

under refrigerated conditions (2 to 5°C) conditions for 23 days under ambient (23 to 28°C) conditions. Under conditions which simulate animal dosing (room temperature, exposed to air in hood), the dosage

formulation showed no appreciable loss.

2.1.3 Flutamide:

Identity: Flutamide

R.O.W. ID No.: 1198E

Source: Sigma Chemical Co.

CAS No.: 13311-84-7

Lot No.: 109H0952

Molecular Wt: 276.2 g/mol

Formula:  $C_{11}H_{11}F_3N_2O_3$ 

Structure:

CF<sub>3</sub>

Purity: 99%

Storage:

Test Article: Store at room temperature (~25°C) in sealed amber glass bottles.

Formulation: Store in sealed amber glass bottles at 5°C or -20°C, protected from

light.

Stability:

Test Article: Analyze every 24± 2 weeks to verify stability.

Formulation: Dose formulations (10 mg/mL in corn oil ) are stable for 42 days at 5°C

or -20°C, with -20°C being preferable. Under conditions which

simulate animal dosing (room temperature, exposed to air in hood), the

dosage formulation showed no appreciable loss.

2.1.4 **Ethinyl Estradiol** 

> Identity: Ethinyl estradiol

R.O.W. ID No.: 1318B

Source: Sig ma Chemical Co.

CAS No.: 57-63-6

45H0716 Lot No.:

Molecular Wt: 296.44

Formula:

Structure:

Purity: 99.7%

Storage:

Test Article: Store in sealed amber glass bottles away from light at ambient

temperatures (23 to 28°C) under inert headspace.

Formulation: Store in sealed amber glass bottles away from light and refrigerated

(~5°C).

Stability:

Test Article: Analyze every 24± 2 weeks to verify stability.

Formulation: Dose formulations (1.0 µg/mL in corn oil ) are stable for up to 14 days

under refrigerated conditions (~5°C). Under conditions which simulate animal dosing (room temperature, exposed to air and light), the dosage

formulation showed no appreciable loss.

# 2.1.5 Certificate of Analysis

Each batch of each test article will be accompanied by a certificate of analysis. The Sponsor will determine for each batch of test article the strength, purity, and composition or other characteristics that appropriately define the test article. A copy of the dose formulation report will be attached (Appendix 2).

#### 2.1.6 Bulk Chemical Samples

Prior to use, two 5 g samples of the bulk of each test article will be collected into a glass bottle with Teflon® coated lids, sealed and stored in the freezer (-20EC) protected from light for possible future reanalysis.

A bulk test article sample of 5 grams will be collected and sent to a NTP subcontractor for purity and stability testing within 30 days of receipt, and thereafter at  $24\pm2$  week intervals. A 35 ml aliquot will be sent within 30 days prior to the start of any study.

#### 2.2 Safety and Handling

The precautions necessary when handling any test article or the prepared formulations of the test substance are based on the Material Safety Data Sheet (MSDS) supplied by the Sponsor. The MSDS will be retained in the study file.

### 2.2.1 Emergency First Aid Procedures

Eye: First check the victim for contact lenses and remove if present. Flush victims eyes with

water or normal saline solution for 20 to 30 minutes while simultaneously calling a hospital or poison control center. Assure adequate flushing. Do not put any ointments, oils, or medication in the victims eyes without specific instructions from a physician. Immediately transport the victim to a hospital even if no symptoms (such as redness or

irritation) develop.

Skin: IMMEDIATELY flood affected skin with water while removing and isolating all

contaminated clothing. Gently wash affected skin areas thoroughly with soap and water.

If symptoms such as inflammation or irritation develop, IMMEDIATELY call a

physician or go to a hospital for treatment.

Inhalation: IMMEDIATELY leave the contaminated area and take deep breaths of fresh air. If

symptoms (such as wheezing, coughing, shortness of breath, or burning in the mouth, throat, or chest) develop, call a physician and be prepared to transport the victim to a

hospital.

Ingestion: If the victim is conscious and not convulsing, give 1 or 2 glasses of water to dilute the

chemical and IMMEDIATELY call a hospital or poison control center. If the victim is

convulsing or unconscious, ensure that the victim's airway is open and lay the victim on his/her side with the head lower than the body. DO NOT INDUCE VOMITING. IMMEDIATELY TRANSPORT THE VICTIM TO A HOSPITAL.

### 2.2.2 Protective Equipment

Eye: Safety glasses/goggles

Gloves: Two pairs of dissimilar protective gloves shall be worn when handling the neat

chemical, and dosing solutions.

Clothing: Minimally, a disposable laboratory suit (e.g. Tyvek ®), bouffant, and shoe

covers shall be worn, as specified in the most current NTP Statement of Work or

the NTP Health and Safety Minimum Requirements.

Respiratory

Protection: A NIOSH-approved chemical cartridge respirator with an organic vapor, acid

gas and high-efficiency particulate filter cartridge. Use only in well ventilated

areas.

# 2.2.3 Spills and Containment

The Health and Safety Officer shall be informed in the event of any spillage. If the spillage is containable (at the discretion of the Health and Safety Officer) the following steps shall be taken:

- 1. A HAZORB® Chemical Spill Kit will be used.
- 2. Place HAZORB® control pillows around the spill area.
- 3. Place additional pillow over spill and allow absorption to occur.
- 4. Dispose of all absorbed material as hazardous waste.

If the spillage is not containable (at the discretion of the Health and Safety Officer), self contained breathing apparatus will be used.

# 2.2.4 Decontamination of Laboratory Equipment

Computer Terminal/ Whenever feasible, a protective covering (*e.g.*, plastic wrap) shall be placed over the keyboard when in use. Before removing gener.

be placed over the keyboard when in use. Before removing general laboratory equipment (*i.e.*, lab carts, portable hoods and balances) from animal dosing rooms and/or chemical preparation areas, clean work

surfaces with a 1% T.B.Q (quaternary ammonium) solution.

# 2.2.5 Disposal Procedures

Waste Disposal: Securely package and label, in double bags, all waste material. All

potentially contaminated material (*i.e.*, carcasses, bedding, soiled disposable clothing) shall be disposed of by incineration in a chemical incinerator equipped with an after burner and scrubber in a manner

consistent with federal (EPA), state, and local regulations.

### 2.3 Dose Formulation and Analysis

The quantity of Vinclozolin and Methoxychlor, Ethinyl estradiol, and Flutamide to prepare a solution will be accurately weighed into a volumetric flask. The vehicle (corn oil) will be added to the required volume and the solution stirred for at least 10 minutes to insure complete dissolution. The formulation for each group will be dispensed into daily aliquots which will be stored in glass bottles with Teflon® coated lids protected from light at 2-9EC. Each solution will be stirred prior to dosing.

Each formulation will be labeled with the TherImmune No., R.O.W. ID No., Group, Dose Level, Vehicle, Mix Number, Preparation Date, Storage conditions, Study and color coded by group. The tray used to hold the daily aliquots will be labeled with the TherImmune Study No., Task No., R.O.W. ID No., Test article Name, Group, Dose Level, Vehicle, Mix Number, Preparation Date, Expiration Date, Storage conditions, Study and Group Color Code (see SOP No 506.0 Storage Sampling, and Labeling of Control and Test Diets and Mixtures and SOP No.121.0 Color-Coding for Study Identification and Dose Groups).

Every time a new mix or batch of chemical is prepared three (3) 35 ml archival samples of each dose level of test article formulation will be collected and stored at TherImmune in glass bottles with Teflon® coated lids protected from light in the refrigerator. One set of samples from each dose level will be forwarded on ice packs to NTP analytical chemistry subcontractor for dose concentration analysis at the following times: initial, mid, and final formulations and at other periods specified by the Sponsor and communicated to the Study Director. If the formulation are suspensions, three archival samples (35 ml) will be collected from the top, mid, and bottom of the low and high dose formulations in glass bottles with Teflon® coated lids. A 5 g sample of bulk test article will be forward to the NTP analytical chemistry subcontractor for stability analysis prior the initial mix and thereafter at  $24 \pm 2$  week intervals. Archival samples which are not selected for analysis will be discarded as hazardous waste as following requirements in Section 2.2.6 after at least ninety days following preparation.

# 3.0 EXPERIMENTAL DESIGN

#### 3.1 Test System

Species: Sprague-Dawley Crl: CD® (SD) IGS BR

Rationale: The Sprague-Dawley rat was selected as the test system due to its established

quality as a breeder and the availability of historical toxicologic data for

reference.

Supplier: Charles River Breeding Laboratories. (Portage, Michigan or Raleigh North

Carolina)

Number/Sex: Thirty time-mated females plus 5 extras will arrive on gestation day (GD) 7-10

(day of mating = GD 0)

Age (at study initiation): Approximately 12 weeks

#### 3.2 Animal Husbandry

All laboratory animal care will be in accordance with the <u>Guide for the Care and Use of Laboratory</u> Animals, TherImmune Standard Operating Procedures, and applicable FDA regulations.

Acclimation period: At least 7 days

#### NTP/NIEHS

Animal housing

during acclimation: 1 /cage

Lighting: 14/10 hour light/dark cycle

Temperature:  $22\pm2^{\circ}$ C

Relative Humidity: 40-50%

Observations: Twice daily observations for general health and availability of adequate

food and water.

Cage changes: At least twice a week, unless the animals are individually housed in 19"

x 10½" x 8" (group-housed) cages which may be changed once a week.

Feeder/bottle changes: At least once per week

Procedure for Individual

Animal Identification: All animals will be uniquely identified by tail tattoo and by cage cards.

Housing Requirements:

Cage Type: Polycarbonate

Cage Measurement: 19" x 10 1/2" x 8" (group housed)

9" x 8 1/2" x 8" (single housed) (20x25x47cm)

Bedding Material: "Sani Chip" Hardwood Laboratory Bedding (P.J. Murphy Forest

Products Corp., Montville, N.J.).

All bedding will be autoclaved prior to use.

Feed: Teklad Certified Rodent Diet 7012C

Frequency: ad libitum

Analysis: The feed is analyzed for nutrients, aflatoxins, nitrosamines, heavy

metals, chlorinated hydrocarbons, organophosphates, PCB's, nitrates, nitrites, BHA, BHT, total bacterial plates, coliforms, E. coli and

Salmonella by the vendor prior to release.

Water: Filtered tap water

Frequency: ad libitum

Analysis A water quality sample is analyzed for total dissolved solids, heavy

metals, chlorinated hydrocarbons, organophosphates, nitrates, nitrites, microbiological content, and total trihalomethanes at least semi-annually to conform with the Safe Drinking Water Act. None of the contaminants are expected to be at levels sufficient to interfere with the

study.

Health Screening Requirements: Prior to initiation of the study one female will be sent to

AnMed/Biosafe Laboratories (Rockville, MD) for serological tests:

Pneumonia Virus of Mice (PVM) Respiratory Enteric Orphan III (REO3) Toolan's H-1 (parvovirus) (TH1) Encephalomyelitis (GD7)
Sialodacryoadenitis Virus (coronavirus)(SDAV/RCV)
Sendai (SEN)
Mycoplasma Pulmonis (MYCO)
Lymphocytic Choriomeningitis (LCM)
Kilham's rat Virus/Rat Orphan Parvovirus (KRV/rOPV)

# 4.0 STUDY DESIGN

#### 4.1 General Study Design

Thirty time-mated females will be used on study and will produce juvenile animals. Ninety juvenile males will be assigned on a male cohort study and ninety juvenile females will be assigned on a female cohort study.

Definition: Gestation Day 0 (GD 0) = Day of mating (Sperm +)Postnatal Day 0 (PND 0) = Day of Delivery

# 4.1.1 Mortality

Any animals found dead or killed *in extremis* on the study will be subject to necropsy. The following tissues will be retained and placed in Bouin's then transferred into 70% ethanol within 24-48 hours:

liver gross lesions
kidneys pituitary
thymus brain
adrenals stomach
spleen thyroid/parathyroids
both testes and epididymis (male)

prostate (ventral and dorso-lateral lobes) (male) seminal vesicles/coagulating glands (male) vagina/uterus/cervix (female) ovaries (female)

Histopathology of the unscheduled deaths/sacrifices will be performed at the discretion of the Sponsor.

# 4.1.2 Necropsy

All necropsies are performed according to TherImmune Standard Operating Procedures.

#### 4.1.3 Histopathology

Histopathological examination of fixed tissues for animals found dead or killed *in extremis* will not be conducted unless indicated by a protocol amendment. Tissues will be transferred to Pathology Associates International (PAI) located in Frederick, MD under subcontract to TherImmune. If pathology is conducted, the findings will be incorporated in the final report.

### 4.2 Juvenile Animal Production

All dams will be housed individually from the day of receiving to the day of euthanasia. Thirty females will be allowed to deliver litters to be used on study. Litters will be culled up to 8-10 pups on PND 4 and approximately equal numbers of male and female pups will be kept in each litter. On PND 21, all male and female pups will be separated from dams and ninety males will be assigned to the juvenile male

study and ninety females will be assigned to the juvenile female study. Selection will be made by weighing all pups and selecting the 90 animals of each sex that are most similar in weight. All extra animals will be removed from study and discarded without necropsy on the day of separation.

#### 4.2.3 Allocation

Thirty time-mated females plus 5 extras will be ordered from Charles River and arrived on gestation day 7-10 (day of mating = gestation day 0).

#### 4.2.4 Treatment

There is no treatment for time-mated females.

# 4.2.5 Measurements

#### **Dams**

Observations for mortality

and clinical signs: Twice daily

Body Weight: At littering

At PND 21

# **Pup Observations**

The following pup observations will be made for the  $F_1$  pups

PND 0: Number of live pups

Number of dead pups Number of males

Total body weight of males

Number of females

Total body weight of females

PND 4: Litters will be culled to 8-10 pups (approximately equal numbers of male and

female pups, if possible). Pups not selected will be discarded without necropsy.

Body weights: Weekly

Individual pup weights will be collected on PND 21 and recorded to the nearest 0.1

gram.

# 4.2.6 Disposition of Offspring and Dams

Dams: Discarded without necropsy on PND 21 following the terminal body weight

collection.

Pups: Animals that are not selected for juvenile male or female cohort studies will be

discarded without necropsy on PND 21.

### 4.3 Juvenile Male Cohort Study:

# 4.3.1 Number of Animals and dose levels:

Group No.	Test Material	Dose Level	Number of Males
		(mg/kg/day)	

1	Corn Oil Vehicle	0	15
2	Vinclozolin	10	15
3	Vinclozolin	30	15
4	Vinclozolin	100	15
5	Flutamide	25	15
6	Flutamide	50	15

#### 4.3.2 Allocation

After PND 21 body weight collection, the male pups in each litter will be assigned to treatment groups in a randomized block-fashion based upon weight, with 6 males per block. The 15 blocks range from heaviest to lightest. Each treatment group then gets one pup from each of the 15 blocks.

#### 4.3.3 Treatment

Starting on PND 23, animals will be administered by oral gavage at 2.5 ml/kg/day (using 18gauge gavage needle at 1 inch length with a 2.25 mm ball and a 1 cc glass disposable tuberculin syringe) once daily at 0700-1000 each day and continue through necropsy. Control animals will receive the vehicle, corn oil, only. The formulations will be stirred before and while dosing. Dose volumes will be calculated daily based on same day body weight.

On the day of termination, animals will be dosed between 0700 to 0900.

#### 4.3.4 Measurements

Observations for mortality

and signs of toxicity: Twice daily

Body Weight: PND 23 and daily thereafter, including preputial separation

and termination

Physical Examination: PND 23 and weekly thereafter

At termination

Preputial Separation Observation: Preputial separation will be observed on all males daily

starting on PND 23. A partial separation, complete separation, and persistent thread of tissue between the gland and prepuce will be recorded. However, the day of complete separation is the endpoint used in the analysis for the age of preputial

separation.

### 4.3.5 Termination

Schedule: On PND 53-54

Groups: The necropsy will examine all treated males from each dose group.

Procedures: Care must be taken to remove mesenteric fat from the sex accessory glands with

small scissors such that the fluids are retained. Once free from the fat and adnexa, the weight with fluid is recorded. The seminal vesicle with coagulating gland is then placed on a paper towel, pressed so that the fluid is exuded, gently

blotted dry, and reweighed.

Small tissues such as the adrenals and pituitary, as well as tissues that contain fluid, will be weighed immediately to prevent partial drying prior to weighing.

Organ Weights: The following organs will be weighed and recorded to the nearest 0.1 mg.

adrenal glands (paired)

epididymis (right and paired weights)

kidneys (paired)

levator ani plus bulbocavernosus

liver pituitary

seminal vesicle and coagulating gland with and without fluid

testes (paired) thyroid/parathyroids

ventral and dorsal-lateral prostate, separately

# Tissue Preservation:

After weighing, the following tissues will be placed in Bouin's and transferred into 70% ethanol within 24-48 hours:

adrenal glands (paired)

epididymides kidneys (paired)

liver pituitary testes

thyroid/parathyroids (with attached portion of the trachea)

gross lesions

Histopathology: The thyroid, testes, and epididymides from all males/group will be embedded in

paraffin, sectioned, stained with hematoxylin and eosin, and examined microscopically. All gross lesions from all males/group will be embedded in paraffin, sectioned, and examined microscopically by the study pathologist.

#### 4.4 Juvenile Female Cohort:

#### 4.4.1 Number of Animals and dose levels

Group No.	Test Material	Dose Level (mg/kg/day)	Number of Females
1	Corn Oil Vehicle	0	15
2	Methoxychlor	12.5	15
3	Methoxychlor	25	15
4	Methoxychlor	50	15
5	Ethinyl Estradiol	0.0025	15
6	Ethinyl Estradiol	0.005	15

#### 4.4.2 Allocation

After PND 21 body weight collection, the female pups in each litter will be assigned to treatment groups in a randomized-block-fashion based upon weight with 6 females per block. The 15 blocks range from heaviest to lightest. Each treatment group then gets one pup from each of the 15 blocks.

#### 4.4.3 Treatment

Starting on PND 22, animals will be administered by oral gavage at 2.5 ml/kg/day day (using 18gauge gavage needle at 1 inch length with a 2.25 mm ball and a 1 cc glass disposable tuberculin syringe), once daily at 0700-1000 each day, and continue through necropsy. Control animals will receive the vehicle, corn oil, only. The formulations will be stirred before and while dosing. Dose volumes will be calculated daily based on same day body weight.

On the day of termination, animals need to be dosed during 0700 to 0900.

# 4.4.4 Measurements

Observations for mortality

and signs of toxicity: Twice daily

Body Weight: PND 22 and daily thereafter

Physical Examination: PND 22 and weekly thereafter

At termination

Vaginal Opening Observation: Vaginal opening will be observed on all females daily starting

> on PND 22. The appearance of a small pin hole, a vaginal thread, and complete vaginal opening are recorded. However, the day of complete vaginal opening is the endpoint used in the analysis for the age of vaginal opening. If vaginal opening does not occur by PND 42 then PND 43 may be used to determine the mean for the age at vaginal opening. In this case, the number of females that did not reach vaginal opening

by necropsy within each treatment group should also be

included in the data summary.

Estrous Cyclicity: Beginning on the day of vaginal opening and continuing

through day of necropsy, daily vaginal smears are obtained and evaluated under a low-power light microscope for the presence of leukocytes, nucleated epithelial cells, or cornified epithelial cells to determine the age of the first vaginal cycle and/or any effects on estrous cyclicity. Extended estrus shall be defined as exhibiting cornified cells with no leukocytes for 3 or more days and extended diestrus as the presence of

leukocytes for 4 or more days.

#### 4.4.4 Termination

Schedule: On PND 42-43

Groups: The necropsy will examine all treated females from each dose group.

Procedures: Care must be taken to remove mesenteric fat from the uterine horns and to avoid

damaging the uterus so that the uterine fluid is retained. The uterus plus cervix are separated from the vagina and the weight of the uterus with fluid is recorded. The uterus is then placed on a paper towel, slit to allow the fluid contents to leak

out, gently blotted dry and reweighed.

Small tissues such as the adrenals and pituitary, as well as tissues that contain fluid, will be weighed immediately to prevent partial drying prior to weighing.

Organ Weights: The following tissues will be weighed and recorded to the nearest 0.1 mg.

adrenal glands (paired)

kidneys (paired)

liver

ovaries (paired)

pituitary

thyroid/parathyroids

uterus and cervix with and without fluid

### Tissue Preservation:

After weighing, the following tissues will be placed in Bouin's and transferred

into 70% ethanol within 24-48 hours:

adrenal glands (paired)

kidneys (paired)

liver

ovaries (paired)

pituitary

thyroid/parathyroids (attached with the portion of the trachea)

uterus and cervix gross lesions

Histopathology: The ovaries, uterus and cervix, and gross lesions from all females/group will be

embedded in paraffin, sectioned, stained with hematoxylin and eosin, and

examined microscopically by the study pathologist.

#### 5.0 PROPOSED STATISTICAL ANALYSES

All raw data will be sent in Excel spreadsheet (soft copy) to the Project Officer at the same time that the data are sent to the statistical support group.

Statistical analyses of the following will be performed:

Data from the main study will be analyzed by a statistical support group under contract to NTP/NIEHS, RTP, NC. A statistical analysis report will be submitted to TherImmune by the contractor for inclusion in the final study report. All data (age and weight at vaginal opening/preputial separation, body and organ weights at necropsy) are analyzed using ANOVA. Organ weights may be analyzed by ANCOVA using the body weight at necropsy as a covariate. When significant treatment effects are observed, treatment means are tested using an appropriate multiple comparison test. Data should be evaluated for heterogeneity of variance by an appropriate statistical test and if present, data should be transformed or analyzed using a suitable non-parametric test.

#### 6.0 REPORTS

The following reports will be submitted:

Draft Study Report

Thirty days after completion of all analysis, all data will be summarized and conclusions on the reproductive toxicity of the test article will be submitted to the Sponsor.

An executive summary will be prepared describing the number and strain of rats observed, the doses used for each chemical tested, and the effects and level of statistical significance for all required endpoints specified in the protocol for the maternal cohort, neonates, uterotrophic cohort and pubertal female/male cohorts. Raw data from each individual animal should be presented in a spreadsheet format. Data summary tables containing the mean, standard error of the mean (SEM), and sample size for each treatment group should be provided for all endpoints. The mean, SEM and CV values for the control data are examined to determine whether they meet acceptable QA criteria for consistency with historical values. Organ weight data may also be presented after covariance adjustment for body weight, but this should not replace presentation of the unadjusted data. In addition, the data tables are accompanied by summary of histological findings with photomicrographs of significant observations.

Final Study Report

The Final Study Report will be submitted to the Sponsor after the submission of the Draft Study Report.

### 7.0 STORAGE OF RECORDS

Upon submission of the final report, all original study records, including all original data sheets; all computer generated data; the original final report; tissues, computer printouts generated in the statistical analysis of data; and copies of the final report will be forwarded to the contracting agency, the NIEHS, Research Triangle Park, North Carolina. Copies of the final study report will also be filed with TherImmune.

#### 8.0 PERSONNEL

Project Officer: Jack Bishop, Ph.D. (NTP)

Study Director:Gary W. Wolfe, Ph.D., D.A.B.TReproductive Toxicologist:Larissa B Nehrebeckyj, B.S.Technical Supervisor:Roland Naawu, M.S., LATG

Health and Safety Officer/

Facility Manager: Robert Blackford, A.A., LATG
Veterinarian: Edward Greenstein, D.V.M, ACLAM

Quality Assurance Officer:Jim Carignan, B.S.Report Manager:Rita Patel, B.S.Dose Preparation Supervisor:Gary Holley, B.S.

# 9.0 SUBCONTRACTORS

Necropsy/Pathology: PAI, Frederick, MD

Serology AnMed/Biosafe, Rockville, MD Clinical Chemistry AniLytics, Inc., Gaithersburg, MD

#### 10.0 REFERENCES

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**Appendix 1: SCHEDULE** 

TO BE ADDED

**Appendix 2: DOSE FORMULATION REPORTS IN STUDY DATA** 

VINCLOZOLIN METHOXYCHLOR FLUTAMIDE ETHINYL ESTRADIOL

**Appendix 3: STANDARD OPERATING PROCEDURES** 

# ADMINISTRATIVE PROCEDURES TABLE OF CONTENTS

Number	<u>Title</u>
101.0	Study Protocol Generation
102.1	Protocol Amendments
103.0	Format of Standard Operating Procedures
105.0	Initiation and Revision of Standard Operating Procedures
105.1	Approval of Health and Safety Standard Operating Procedures
106.0	Review Process of Standard Operating Procedures
107.0	Distribution of Standard Operating Procedures
108.0	Historical SOP File
109.0	Reproduction of Standard Operating Procedures
111.0	Assignment of Study Directors
112.0	Procedures for Manual Data Recording and Correction
112.1	Quality Control Procedures
113.0	Documentation of Protocol and SOP Deviations
114.0	Final Report Format
115.0	Amendment or Revision of Final Reports
116.0	Transfer of Pathology Data, Specimens, and Materials
117.0	Personnel Movement – Second Floor
117.1	Personnel Movement – First Floor
118.0	Archive Procedures
119.0	Maintenance of Personnel Training and Experience Records
120.0	Assignment of TherImmune Research Corporation Study and ID Numbers
121.0	Color-coding for Study Identification and Dose Groups
122.0	Animal Study Notebook
123.0	Regulatory Inspections
124.0	IACUC Procedures
125.0	Storing Study-Related Correspondence

# ANIMAL FACILITIES AND CARE TABLE OF CONTENTS

Number	<u>Title</u>
201.0	Ordering Animals and Materials/Supplies
202.0	Animal Receipt, Housing and Quarantine for Rodents
202.1	Animal Receipt, Housing and Quarantine for Non-Rodents
204.0	Animal Housing and Room Set Up for Rodents
205.0	Collection of Temperature and/or Humidity in the Artemis Data Collection System
207.0	Animal Husbandry and Observations
207.1	Operation of Bacharach Sling Psychrometer
207.2	Animal Husbandry and Observations for Non-Rodents
207.4	Environmental Enrichment for Laboratory Animals
207.5	Collection of Animal Husbandry Data in the Artemis Data Collection System
208.0	Whole Animal Health Screen
209.0	Receipt and Storage of Animal Feed and Bedding
211.0	Water and Feed Analysis
212.0	Rodent Serology- Plasma Collection
213.0	Rack Rotation (Rodent)
214.0	Animal Identification by Tattooing for Rodents
214.1	Animal Identification by Cage Cards
214.2	Animal Identification by Cage Cards for Non-Rodents
214.3	Correlation of Supplier Number to TherImmune Research Corporation Animal ID Number for Non-Rodents
214.4	Animal Identification by Ear Tag for Rodents
214.5	Implanting and Mapping with Electronic Laboratory Animal Mapping Systems
214.6	Animal Identification and Data Collection with Electronic Laboratory Animal Mapping Systems
215.0	Assignment of Unique Animal Identification Numbers
216.0	Clinical Pathology Sampling via Orbital Sinus - Rodents
216.1	Clinical Pathology Sampling via Jugular Vein - Canine
216.2	Clinical Pathology Sampling via Medial Auricular Artery - Rabbits
216.3	Clinical Pathology Sampling via Femoral Vein - Non-Human Primate
216.4	Clinical Pathology Sampling via Abdominal Aorta/Vena Cava - Rodents
216.5	Clinical Pathology Sampling via Jugular Vein – Rodents

216.6	Clinical Pathology Sampling via Anterior Vena Cava - Swine
217.0	Preparation of Blood Smears - Differential/Morphology and Reticulocytes
217.1	Preparation of Bone Marrow Smears from Rodents
218.0	Microisolator Cage Changing
220.0	Sanitation of Animal Transport Vehicle
221.0	Intradermal Testing for Mammalian Tuberculosis
222.0	Recording General Information for Studies
223.0	Urinalysis Using CHEMSTRIP® Urine Test Strips
224.0	Collection of Temperature and/or Humidity in the Artemis Data Collection System

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301.1	Additional Clinical Observations/Veterinary Services
302.2	Collection of Body Weights in the Artemis Data Collection System
302.3	Collection of Food and Water Consumption Data in the Artemis Data Collection System
302.4	Collection of Physical Examinations and/or Cageside Observations using the Artemis
	Data Collection System
302.5	Procedures for "Invalid Results" in Artemis
302.6	Procedures for Out of Range Data in the Artemis Data Collection System
303.2	Back-Entering Data Into Artemis Data Collection System
304.0	Feed and Water Dosing
305.0	Administration of Test Article by Oral Gavage - Rodents

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400.1	Confirmation of Copulation and Separation for Reproductive Studies
400.2	Confirmation of Mating and Separation of Males and Females in the Artemis Data Collection System
400.3	Mating in the Artemis Data Collection Data System
401.0	Cohabitation Procedures for Rodent Reproductive Studies
401.1	Cohabitation Procedures for Males and Females in the Artemis Data Collection System
402.0	Preparation and Evaluation of Vaginal Smears for Determination of Estrous Cycles
402.1	Determination of Estrous Cycle in the Artemis Data Collection System
403.0	Parturition and Neonatal Observations
403.1	Littering in Progress Documentation in the Artemis Data Collection System
403.2	Parturition and Neonatal Observation in the Artemis Data Collection System
403.3	Culling Pups in the Artemis data Collection System
403.4	Pup Identification in the Artemis Data Collection System
405.0	Postweaning Sexual Development Evaluation in Rodents
405.1	Recording Postweaning Sexual Development Data in the Artemis Data Collection System
407.0	Random Selection of Weanling Rodents
407.1	Random Selection of Weanling Rodents in the Artemis Data Collection System
414.0	Preparation of Buffers
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501.0	Control and Test Article Receipt and Use
502.0	Maintenance of Material Safety Data Sheet (MSDS)
504.0	Receipt and Labeling of Reagents and Solutions
505.0	Verification of Quality of Reagents Upon Receipt from Commercial Sources
506.0	Storage, Sampling, and Labeling of Control, Test Diets and Mixtures
506.1	Shipping of Surplus Test Articles, Control Articles and/or Samples
507.0	Dose Formulation Calculations
509.0	Formulation File
510.1	Corn Oil Peroxide Analysis - NIEHS/NTP
511.0	Preparation of Test FormulationsSolutions/Suspensions, Test Diet, and Dosed Water
513.0	Dispensing of Test Formulations
514.0	Cleaning Procedures in Formulations
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603.0	Data Security
607.0	Working Group Definitions and Procedures in the Artemis Data Collection System
608.0	Logon/Logoff in the Artemis Data Collection System
609.0	Creating a Stock Study in the Artemis Data Collection System
610.0	Creating a Target Study in the Artemis Data Collection System
611.0	Control and Documentation of Glossary Changes
612.0	Printing Data Report in Artemis
612.1	Creating New Versions of the Report Document in Artemis
612.2	Printing Reports in the Artemis II Table Production Module
612.3	Printing the Work Schedule Report in Artemis
613.0	Inserting Histopathology Report into the Main Report Document
614.0	Report Format Specification in Artemis
615.0	Inserting Outside Contributions in to the Main Report Document
616.0	Running and Reporting Statistics in Artemis
617.2	Updating Glossaries in the Artemis Pathology Module
617.1	Creating Glossaries in the Artemis Pathology Module
617.3	Collection of Necropsy Data in the Artemis Pathology Module
617.4	Amending Necropsy Data in the Artemis Pathology Module

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701.0	Tunnel Washer Operation and Maintenance
702.0	Chemical Fume Hood Operation
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704.0	Thermometer Calibration
705.0	Calibration, Operation and Certification of Mettler Balances
705.1	Calibration, Operation and Certification of Detecto Electronic Ba lances
706.0	Documenting Refrigerator, Freezer, and Incubator Temperatures
707.0	CESCO 407 Series Rack Washer Operation and Maintenance
711.0	Edstrom/SE Lab Automatic Watering Maintenance
712.0	Boiler Operation and Maintenance
713.0	Emergency Generator Testing
714.0	Verification of Air Direction/Number of Changes
715.0	Heating, Ventilation, and Air Conditioning Equipment Maintenance
716.0	Facility Access and Security
717.0	Pest Control
719.0	Manual Disinfection Procedures
720.0	Taylor Hygrometer Use
721.0	Waste Management System Operation
721.1	Waste Management
722.0	Maintenance of Light Cycles
724.1	Operation of the Amsco Autoclave
725.0	Operation of Stir Plates
726.0	Operation of the REES Environmental and Security System
727.0	Autoclave Sterility Testing
728.0	Operation, Maintenance, and Calibration of Mitutoyo Calipers
729.0	Operation and Cleaning of the Lab-Line® Ultrasonic Bath and the Bransonic®
	Ultrasonic Cleaner
730.0	Operation and Cleaning of the Dynac® and Adams® Centrifuges
731.0	Operation and Calibration of Micropipettes
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# QUALITY ASSURANCE UNIT TABLE OF CONTENTS

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809.0	In-Life Data Audits for NIEHS Studies
812.0	Audit Procedures for Reports
813.0	Post audit Reviews of Final Reports
814.0	Quality Assurance Statements
815.0	Responses to Quality Assurance Inspection/Audit Status Reports
819.0	Quality Assurance Study Files
820.0	Facility Inspection
822.0	Distribution of Inspection and Audit Status Reports
823.0	Quality Assurance Monthly Status Reports
824.0	Rejection Criteria for Data and Final Reports
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900.0	Rodent Necropsy - NIEHS/NTP Studies
901.0	Euthanasia of Rodents Using Inhalants and Physical Methods
901.1	Euthanasia Using Sodium Pentobarbital
903.2	Preparation of Miscellaneous solutions
904.0	Anesthesia by CO <sub>2</sub> /O <sub>2</sub> Inhalation
906.0	Small Animal Necropsy

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1002.0	Respiratory Protection Program
1003.0	Personal Protective Equipment
1005.0	Materials Control – Receiving/Warehouse Areas, Active Usage Areas, Waste Storage
	Area, Emissions Systems, and Waste Disposal
1006.0	Facility Control
1007.0	Emergency Response
1008.0	Waste Disposal
1009.0	Medical Surveillance Program
1010.0	Disaster Plan

# References

Visitors Access to Test Area 117.0, CHP, RPP

Employee Training 119.0, 1001.0, CHP, RPP

Medical Surveillance 1001.0, CHP, RPP

Eye Protection 117.0, 1003.0, CHP

Personal Protective Equipment 1003.0, CHP, RPP

General Housekeeping Practices 1005.0, CHP Ventilation System Maintenance 715.0, CHP

Storage, Receipt, Transport and 501.0, 502.0, 504.0

Shipping of Study Materials 506.0, 506.1,1005.0, CHP

Spill Clean-Up, Accident and Emergency 1007.0, CHP

Response (including material disasters) and

fires/explosions

Dose Preparation 500 Series

Enter and Exit from Limited Access Areas 117.0, CHP, RPP

Respiratory Protection and Fit RPP

Note: CHP = Chemical Hygiene Plan

RPP = Respiratory Protection Program

# STANDARD OPERATING PROCEDURES FOR HEALTH AND SAFETY TABLE OF CONTENTS

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1100.0	Computer System Disaster Recovery
1101.0	Inspections and Maintenance of Computer System
1102.0	Systems Development Methodology
1103.0	Requirements for Installing Hardware and Operating System
1104.0	Computer Security Issues
1106.0	Setting up a Password in Artemis
1107.0	Version Change Control
1108.0	Software Incident Report (SIR)
1109.0	Client Installation
1109.1	Artemis II Version 4.0.2.3 Client Installation
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